B2 Figure 1 depicts a nucleotide sequence of mouse *Fkh*^{sf} Cdna (SEQ ID NO: 1); translation is predicted to initiate at position 259 and terminate at position 1546.

Please replace the paragraph beginning at page 5, line 7, with the following rewritten paragraph:

B3 Figure 2 depicts the amino acid sequence of mouse Fkh^{sf} (SEQ ID NO: 2).

Please replace the paragraph beginning at page 5, line 9, with the following rewritten paragraph:

B4 Figure 3 depicts a nucleotide sequence of 1735 bp corresponding to human *FKHsf* cDNA (SEQ ID NO: 3; including a 1293 bp coding region); translation is predicted to initiate at position 55 and terminate at position 1348.

Please replace the paragraph beginning at page 5, line 12, with the following rewritten paragraph:

B5 Figure 4 depicts the sequence of a 431 amino acid human FKH^{sf} protein (SEQ ID NO: 4).

Please replace the paragraph beginning at page 34, line 4, with the following rewritten paragraph:

B6 A complementary DNA (cDNA) encoding the complete mouse Fkh^{sf} protein may be obtained by a reverse-transcriptase polymerase chain reaction (RT-PCR) procedure. More specifically, first-strand cDNA is generated by oligo dT priming 5 ug of total RNA from a suitable source (eg., mouse spleen) and extending with reverse transcriptase under standard conditions (eg., Gibco/BRL SuperScript kit). An aliquot of the first-strand cDNA is then subjected to 35 cycles of PCR (94°C for 30 sec, 63°C for 30 sec, 72°C for 2 min) in the presence of the forward and reverse primers (Forward primer: GCAGATCTCC TGACTCTGCC TTC; SEQ ID NO: 5; Reverse primer: GCAGATCTGA CAAGCTGTGT CTG; SEQ ID NO: 6) (0.2 mM final concentration), 60 mM

Tris-HCl, 15 mM ammonium sulfate, 1.5 mM magnesium chloride, 0.2 mM each dNTP and 1 unit of Taq polymerase.

Please replace the paragraph beginning at page 34, line 18, with the following rewritten paragraph:

A human *FKH*^{sf} cDNA encoding the complete FKH^{sf} protein may be obtained by essentially the same procedure as described in Example 2. In particular, starting with total spleen RNA, and utilizing the following oligonucleotide primers (Forward primer: AGCCTGCCCT TGGACAAGGA C; SEQ ID NO: 7; Reverse primer: GCAAGACAGT GGAAACCTCA C; SEQ ID NO: 8), and the same PCR conditions outlined above, except with a 60°C annealing temperature.

Please replace the paragraph beginning at page 35, line 11, with the following rewritten paragraph:

As an example, a 360 bp DNA fragment is amplified from 1st strand cDNA using the following oligos:

DMO5985 (forward): CTACCCACTGCTGGCAAATG (ntd. 825-844 of Figure. 1)
(SEQ ID NO: 9)

DMO6724 (reverse): GAAGGAACTATTGCCATGGCTTC (ntd 1221-1199)
(SEQ ID NO: 10)

Please replace the paragraph beginning at page 35, line 20, with the following rewritten paragraph:

The PCR products are run on an 1.8% agarose gel, transferred to nylon membrane and probed with end-labeled oligonucleotides that are complementary to the region corresponding to the site of the Scurfy-specific 2 bp insertion. Two separate hybridization reactions are performed